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(54) Interferon conjugate and production thereof using recombinant gene.

(57) An interferon conjugates comprising in a single molecule a region exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological activities of interferon- $\gamma$ ; deoxyribonucleotide sequences coding for the interferon conjugate; recombinant DNAs comprising the deoxyribonucleotide sequence and a deoxyribonucleotide sequence coding for a control region for expression of the interferon conjugate wherein the latter sequence is upstream from the former sequence; transformant organisms transformed with the recombinant DNA; and a process for production of the above-mentioned interferon conjugate comprising the steps of culturing the transformant organism to produce the interferon conjugate, and recovering the interferon conjugate.

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INTERFERON CONJUGATE AND PRODUCTION THEREOF  
USING RECOMBINANT GENE

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates to interferon conjugates comprising interferon- $\beta$  and interferon- $\gamma$ , a  
5 process for production of these interferon conjugates, and a gene system including a nucleotide sequence coding for these interferon conjugate, a recombinant DNA comprising the nucleotide sequence, and microorganisms transformed with the recombinant DNA.

10 2. Description of the Related Art

Interferons are proteins exhibiting various biological activities, such as antiviral activity, and therefore, are promising for clinical application.

Interferons are classified into three types,  
15 i.e., interferon- $\alpha$ , - $\beta$ , and - $\gamma$ , depending on their inducers, producer cells or antigenecities thereof, and are different in structure gene coding therefor, properties as protein, and in biological activities (Interferon no Kagaku (Science of Interferon), edited by  
20 Shigeyasu Kobayashi, Kodan Sha).

Interferon- $\beta$  (IFN- $\beta$ ) is a glycoprotein usually produced by fibroblast induced with virus or double-stranded RNA, is stable to treatment at pH 2, and unstable to treatment at 56°C. Gene coding for  
25 interferon- $\beta$  has been already isolated (Taniguchi et al., Proc. Jpn. Acad., 55, Ser. B, 464-468, 1979). The nucleotide sequence of the gene and a corresponding amino acid sequence have been determined, and moreover, a process for the production of interferon- $\beta$  using cDNA thus obtained and E. coli as a host has been developed  
30 (Taniguchi et al., Proc. Natl. Acad. Sci. USA, 77, 5230-5233, 1980; Goeddel et al., Nucleic Acids Res., 8, 4057-4074, 1980; and Derynck et al., Nature, 287,

193-197, 1980).

Interferon- $\gamma$  (IFN- $\gamma$ ) is a glycoprotein usually produced by T lymphocyte induced with a mitogen, and is unstable to treatment at pH 2. A gene coding for  
5 interferon- $\gamma$  has been also isolated, the nucleotide sequence thereof has been determined, and a process for the production of interferon- $\gamma$  using E. coli as a host has been established (Devos et al., Nucleic Acids Res. 10, 2487-2501, 1982; and Gray et al., Nature, 295,  
10 503-508, 1982). Moreover, an amino acid sequence of native interferon- $\gamma$  has been reported (Rinderknecht et al., J. Biol. Chem., 259, 6790-6797, 1984).

Among the interferons- $\alpha$ , - $\beta$ , and - $\gamma$ , interferons- $\alpha$  and - $\beta$  have been known as interferon I,  
15 and it has been suggested that these interferons have a high structural analogy showing a 29% amino acid sequence homology (Taniguchi et al., Gene, 10, 11-15, 1980), and it is believed that they recognize same receptor. Therefore, the coexistence of interferons- $\alpha$  and - $\beta$  provides their activities only in an additive  
20 manner. On the contrary, interferon- $\gamma$ , known as interferon II, has a low amino acid homology with interferon I. It is believed that interferons I and II recognize different receptors (Branca et al., Nature, 294, 768-770, 1981). Therefore, interferons I and II  
25 exhibit different spectra of the anti-cell proliferation effect (Interferon no Kagaku (Science of Interferon), edited by Shigeyasu Kobayashi, Kodan Sha, Japan, 22-68, 1985), and exhibit their activities in a synergistic manner (Czarniecki et al., J. Virol., 49, 490-496, 1984;  
30 Fleishmann Jr. et al., J. IFN. Res., 4, 265-274, 1984; and Japanese Unexamined Patent Publication 59-98019).

Although a mixture of interferons- $\beta$  and - $\gamma$  will show the above-mentioned synergism in vitro, it is  
35 doubtful whether such synergism is established in vivo because there is some possibility that both interferon- $\beta$  and - $\gamma$  would not be simultaneously present at a target

site due to their different in vivo pharmacodynamics.

To resolve the above-mentioned disadvantage provided by the problems of in vivo pharmacodynamics, it would be effective to link interferons- $\beta$  and - $\gamma$  to make  
5 a single polypeptide which could exhibit in vivo the same synergism as shown in vitro by a mixture of interferons- $\beta$  and - $\gamma$ . This means that, since a linked polypeptide molecule exhibits the same synergistically enhanced actions as shown by the parent two interferon  
10 molecules, i.e., - $\beta$  and - $\gamma$ , the activity per a molecule is enhanced in comparison to the activity per molecule of the parent interferons, making it possible to obtain a new type of interferon with a higher activity.

Moreover, by causing a single polypeptide to  
15 exhibit the different kinds of actions originally exhibited separately by interferons- $\beta$  and - $\gamma$ , it would be possible to obtain a new type of interferon having a broader spectrum of actions.

Some examples are already known wherein two  
20 polypeptides having different actions are linked to make a single polypeptide having two actions (Yournon et al., Nature, 228, 820-824, 1970; Neuberger et al., Nature, 312, 604-608, 1984; and Bulow et al., Biotechnology, 3, 821-823, 1985). Moreover, insulin molecules were linked  
25 to make a stable polypeptide (Shen and Shi-Hsiang, Proc. Natl. Acad. Sci. USA, 81, 4627-4631, 1984). As another example, there was disclosed a linkage of interferon- $\gamma$  and interleukin-2 to make a single polypeptide which exhibits both actions of interferon- $\gamma$  and interleukin-2  
30 (Japanese Unexamined Patent Publication No. 60-241890).

However, so far there has been no attempt to link interferon- $\beta$  with interferon- $\gamma$  to prepare a single polypeptide having at the same time both a broader spectrum of actions and higher activities.

#### 35 SUMMARY OF THE INVENTION

Therefore, the present invention relates to interferon conjugates comprising interferon- $\beta$  and

interferon- $\gamma$ , which have a broader spectrum of biological actions, such as antiviral action, anti-cell proliferation action, and the like, that were originally carried separately by separate polypeptides, i.e.,  
5 interferon- $\beta$  and interferon- $\gamma$ , and which exhibits in vivo synergistically enhanced activities as shown in vivo by a mixture of interferons- $\beta$  and - $\gamma$ .

More specifically, the present invention provides an interferon conjugate comprising in a single molecule  
10 a region exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological activities of interferon- $\gamma$ .

The present invention also provides a deoxyribonucleotide sequence coding for this interferon conjugate.

15 Moreover, the present invention provides a recombinant DNA comprising the above deoxyribonucleotide sequence and a deoxyribonucleotide sequence coding for a control region for expression of the interferon conjugate, wherein the latter sequence is present  
20 upstream from the former sequence.

Moreover, the present invention provides a transformant organism transformed with the recombinant DNA.

Moreover, the present invention provides a process for production of the above-mentioned interferon  
25 conjugate comprising the steps of culturing the transformant organism to produce the interferon conjugate, and recovering the interferon conjugate.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 represents an embodiment of an amino acid  
30 sequence of human mature interferon- $\beta$ ;

Fig. 2 represents an embodiment of an amino acid sequence of human mature interferon- $\gamma$ ;

Fig. 3 represents a structure of plasmid pKM6 for expression of human interferon- $\beta$ ;

35 Fig. 4 represents a structure of plasmid p6hu $\gamma$ -N1 for expression of human interferon- $\gamma$ ;

Fig. 5 represents a structure of plasmid pKM6-Cxho

into which an XhoI site has been introduced at the 3'-terminus of the human interferon- $\beta$  structural gene;

Fig. 6 represents a structure of plasmid p6huyN1-CKpn into which a KpnI site has been introduced at the 3'-terminus of the human interferon- $\gamma$  structural gene;

Fig. 7 represents a structure of plasmid p6huyN1ABS-NHin into which a HindPI site has been introduced to take out a human interferon- $\gamma$  structural gene;

Fig. 8 schematically represents a process for construction of a plasmid for expression of an interferon- $\gamma$ - $\beta$  conjugate;

Fig. 9 schematically represents a process for construction of a plasmid for expression of an interferon- $\beta$ - $\gamma$  conjugate;

Fig. 10 schematically represents a process for construction of a plasmid for expression of an interferon- $\gamma$ - $\beta$  conjugate;

Fig. 11 represents a nucleotide sequence coding for an amino acid sequence of a spacer peptide;

Fig. 12 schematically represents a process for construction of a plasmid for expression of an interferon- $\beta$ - $\gamma$  conjugate;

Fig. 13 schematically represents a process for construction of a plasmid for expression of an interferon- $\gamma$ - $\beta$  conjugate in animal cells; and

Fig. 14 schematically represents a process for construction of a plasmid for expression of an interferon- $\gamma$ - $\beta$  conjugate in animal cells.

In Figures 3 to 10 and 12, the numbers 1, 2, and 3 represent a human interferon- $\beta$  structural gene, a human interferon- $\gamma$  structural gene, and a non-coding region of human interferon- $\gamma$  cDNA, respectively.

#### DESCRIPTION OF THE PREFERRED EMBODIMENT

The interferon conjugate of the present invention comprises a region exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological

activities of interferon- $\gamma$  linked directly or via a  
spacer. According to the present invention, the terms  
"interferon- $\beta$ " and "interferon- $\gamma$ " include any proteins  
having activities specific to interferon- $\beta$  and  
5 interferon- $\gamma$ , respectively. For example, for  
interferon- $\gamma$ , a modified interferon- $\gamma$  wherein its  
polypeptide has been extended by three amino acid  
residues at N-terminal (Gray et al., Nature, 295,  
503-508, 1982), and a modified interferon- $\gamma$  wherein a  
10 part of the C-terminal of the polypeptide thereof has  
been deleted (Rose et al., Biochem. J., 215, 273, 1983),  
are known. These modified interferon- $\gamma$  fall under the  
scope of the present invention provided that they  
maintain interferon activities. Moreover, modified  
15 interferon- $\gamma$  wherein some amino acid residues have been  
replaced by other amino acid residues are disclosed  
(Japanese Unexamined Patent Publications No. 59-93093  
and No. 59-167596). These modified interferon- $\gamma$  also  
fall under the scope of the present invention provided  
20 that they maintain interferon activities. Interferon- $\beta$   
also can be modified by the addition, deletion or  
replacement of one or more amino acid residues as  
described for interferon- $\gamma$ . Preferably, interferon- $\beta$   
comprises a polypeptide having the amino acid sequence  
25 shown in Fig. 1, and interferon- $\gamma$  comprises a polypeptide  
having the amino acid sequence shown in Fig. 2.

Juxtaposition order of interferons- $\beta$  and - $\gamma$  in the  
present interferon conjugate is not limited. That is,  
the interferon conjugate can contain as its N-terminal  
30 half a polypeptide of interferon- $\beta$  and as its C-terminal  
half a polypeptide of interferon- $\gamma$ , and vice versa.

In junction region in an interferon conjugate, a  
polypeptide of interferon- $\beta$  and that of interferon- $\gamma$  can  
be linked directly, or indirectly via a spacer, such as  
35 a peptide. The spacer peptide is preferably a peptide  
comprising many hydrophilic amino acids, as shown in an  
example for an enzyme wherein subunits of  $\beta$ -galactosidase

are linked via a spacer peptide (Kushinke et al, EMBO J., 4, 1067-1073, 1985). Moreover, a polypeptide which links domains of a naturally occurring protein can be used as a spacer peptide of the present invention. A  
5 spacer peptide usually consists of up to 50 amino acids. A preferred spacer peptide is, for example, a peptide known as a switch-peptide of immunoglobulin molecule, and also a peptide having the amino acid sequence:  
Thr-Gln-Leu-Gly-Gln-Pro-Lys-Ala-Ala-Lys-Ser-Val-Thr.

10 In the present invention, a structure comprising a polypeptide of interferon- $\beta$  and a polypeptide of interferon- $\gamma$  is designated as "interferon conjugate". More specifically, a structure comprising as its  
N-terminal half a polypeptide of interferon- $\beta$  and as  
15 its C-terminal half a polypeptide of interferon- $\gamma$  is designated as "interferon- $\beta\cdot\gamma$  conjugate" (IFN- $\beta\cdot\gamma$ ); and a structure comprising as its N-terminal half a polypeptide of interferon- $\gamma$ , and at its C-terminal  
half, a polypeptide of interferon- $\beta$ , is designated as  
20 "interferon- $\gamma\cdot\beta$  conjugate" (IFN- $\gamma\cdot\beta$ ). On the other hand, a structure comprising as its N-terminal half a polypeptide of interferon- $\beta$  and as its C-terminal half a polypeptide of interferon- $\gamma$  wherein both polypeptides  
are linked via a spacer is designated as an "interferon-  
25  $\beta\gamma$  conjugate" (IFN- $\beta\gamma$ ); and a structure comprising as its N-terminal half a polypeptide of interferon- $\gamma$  and as its C-terminal half a polypeptide of interferon- $\beta$   
wherein both polypeptides are linked via a spacer is  
designated as an "interferon- $\gamma\beta$  conjugate" (IFN- $\gamma\beta$ ).  
30 As processes of the production of an interferon conjugate, there are a chemical synthesis wherein amino acids are coupled in a predetermined order, and genetic engineering process wherein a DNA coding for a target  
polypeptide is designed and constructed and the target  
35 polypeptide is expressed in appropriate transformed cells. In the present invention, preferably the genetic engineering is used because it enables an easy production



of a target polypeptide, though the means of obtaining a target polypeptide is not limited to genetic engineering.

In the use of genetic engineering for the production of an interferon conjugate, a DNA fragment comprising  
5 a nucleotide sequence coding for a polypeptide of interferon- $\beta$  and a nucleotide sequence coding for a polypeptide of interferon- $\beta$ , wherein these nucleotide sequences are directly linked or indirectly linked via a nucleotide sequence coding for a spacer peptide, are  
10 prepared, and this DNA fragment is linked to appropriate control regions for expression of the target polypeptide, and the coding sequence is expressed in appropriate host cells.

Any nucleotide sequence coding for an interferon  
15 conjugate can be used. That is, if more than one codon is present for an amino acid, any codon which codes for the amino acid can be used. Preferably, the same nucleotide sequences as those of the cDNAs of interferons- $\beta$  and - $\gamma$  respectively are used (Taniguchi  
20 et al., *Gene*, 10, 11-15, 1980; and Devos et al., *Nucleic Acids Res.*, 10, 2487-2501, 1982).

To obtain a DNA fragment having a nucleotide sequence coding for an interferon conjugate, chemical synthesis of the DNA, isolation and linking of genes  
25 coding for interferons- $\beta$  and - $\gamma$ , and a combination of such means can be used. The chemical synthesis of a DNA fragment having a desired nucleotide sequence can be achieved according to a conventional procedure (Edge et al., *Nature*, 292, 756-762, 1981; and Tanaka et al.,  
30 *Nucleic Acids Res.*, 11, 1707-1723, 1983). DNA fragment coding for interferon- $\beta$  or - $\gamma$  can be derived from chromosomal DNA, or preferably, prepared as cDNA. cDNA can be isolated according to a known procedure (Taniguchi et al., *Proc. Jpn. Acad.*, 55, Ser. B, 464,  
35 1979; Goeddel et al., *Nucleic Acids Res.*, 8, 4057-4074, 1980; Derynck et al., *Nature*, 287, 193-197, 1980; Devos et al., *Nucleic Acids Res.* 10, 2487-2501, 1982; and Gray

et al., Nature, 295, 503-508, 1982). Alternatively, the desired DNA fragment can be obtained by screening a cDNA library prepared according to a known procedure (Okayama et al., Molecular and Cellular Biology, 3, 280, 1983)

5 and using a probe designed from a known nucleotide sequence described in the above-mentioned literature.

To obtain a DNA fragment coding for an interferon conjugate from cDNAs of interferons- $\beta$  and - $\gamma$ , the cDNAs are separately digested with an appropriate restriction  
10 enzyme, and the resulting cDNA fragments are ligated immediately or after blunt-ending with a mung bean nuclease, DNA polymerase I Klenow fragment or T4 DNA polymerase. The cDNA fragments are preferably linked via a synthetic oligonucleotide corresponding to  
15 nucleotides deleted by the restriction enzyme cleavage, to obtain a DNA fragment coding for an interferon conjugate comprising entire polypeptides of interferons- $\beta$  and - $\gamma$ . In such a case, according to one embodiment of the present invention, the synthetic oligonucleotide  
20 consists of a nucleotide sequence coding for a spacer peptide flanked by nucleotide sequences corresponding to nucleotides deleted by the restriction enzyme cleavage, so as to obtain a DNA fragment coding for an entire interferon conjugate comprising polypeptides of  
25 interferons- $\beta$  and - $\gamma$  linked via the spacer peptide. Alternatively, a restriction enzyme site is introduced in cDNAs of interferon- $\beta$  and interferon- $\gamma$  at their 5'-end or 3'-end of the structural gene using synthetic oligonucleotides, and the cDNAs are cleaved with an  
30 appropriate restriction enzyme, and if necessary, after the cDNAs are blunt-ended, the cDNAs are ligated. In any case, one cDNA fragment should be linked in reading frame with another cDNA fragment.

To produce an interferon conjugate using the  
35 above-mentioned DNA construct, animal or plant cells, yeast or E. coli are used as a host. To express an interferon conjugate, the DNA construct should have a

promoter for starting the transcription, and an SD sequence and ATG codon for starting the translation upstream therefrom. As a promoter, any nucleotide sequence having a promoter activity can be used; for  
5 example, lac promoter, trp promoter, recA promoter, and the like are known. Preferably a strong promoter such as trp promoter is used. The SD sequence is a ribosome RNA binding site and, therefore, essential for translation. In the present invention, any SD sequence  
10 can be used. A control region comprising the above-mentioned promoter and SD sequence thus constructed is added to a DNA fragment coding for an interferon conjugate via an ATG codon to express the interferon conjugate. The addition of the ATG codon can be carried  
15 out using synthetic oligonucleotides (Goeddel et al., Nature, 281, 544-548, 1979). Alternatively, for interferon- $\beta$ , the ATG codon can be exposed according to a known procedure (Taniguchi et al., Proc. Natl. Acad. Sci. USA, 77, 5230-5233, 1980).

20 To introduce the thus-obtained DNA into host cells, a vector DNA is used. Vector DNAs used for an E. coli host include a plasmid DNA such as pBR322 and pSC101, and a phage DNA such as  $\lambda$  phage. Such a DNA vector is linked with the DNA construct for expression of  
25 an interferon conjugate to construct an expression DNA vector, which is then used to transform host cells according to a known procedure (Maniatis et al., "Molecular Cloning", Cold Spring Harbor Laboratory, p250-255, 1982).

30 A transformant such as transformed E. coli is then cultured in an appropriate medium to produce an interferon conjugate. Culturing is preferably carried out in a liquid medium. When trp promoter is used, indoleacrylic acid is added to a culture medium during  
35 culturing to induce the production of an interferon conjugate. If a different promoter is used, an inducer selected in accordance with the particular promoter is

preferably used, to enhance the production of an interferon conjugate.

The interferon conjugate producer such as the thus-cultured E. coli cells are then disrupted according  
5 to a conventional cell disruption procedure ("Tanpakushitsu-Koso no Kisojikken" (Fundamental Experiment of Protein and Enzyme), edited by Takekazu Horio and Jinpei Yamashita, Nankodo, 1981, 3-7), for  
10 example, by treatment with a enzyme, sonication, grinding, or treatment under pressure, to obtain a crude extract containing an interferon conjugate.

The above-mentioned cell disruption procedure can be combined with treatment with guanidine hydrochloride or urea to improve the extraction efficiency (Davis et  
15 al., Gene, 21, 273-284, 1983).

The thus-obtained crude extract can be further purified according to a known procedure ("Tanpakushitsu-Koso no Kisojikken" (Fundamental Experiment of Protein and Enzyme), edited by Takekazu  
20 Horio and Jinpei Yamashita, Nankodo, 1981, 18-382), for example, by salting out, ultrafiltration, ion exchange, gel filtration, affinity chromatography, electrophoresis, or a combination thereof, to prepare a highly purified interferon conjugate preparation.

25 To express an interferon conjugate in animal cells, it is necessary that a nucleotide sequence coding for the interferon conjugate be present under the control of a promoter effective in the animal cells. Promoters effective in animal cells include, for example, SV40  
30 early promoter, SV40 late promoter, a promoter of a HB virus gene, MMTV promoter, a promoter of a thymidine kinase gene, a promoter of a heat shock protein, and a promoter of an interferon gene. A nucleotide sequence coding for an interferon conjugate is linked downstream  
35 of one of the above-mentioned promoters in the same manner as described for the expression in E. coli. A single promoter or a combination of more than one

promoter can be used. Not, an enhancer sequence of Harbey mouse sarcoma virus 5'LTR or an enhancer sequence of SV40 can be linked upstream of the above-mentioned promoter for a eukaryotic cell. Such enhancers are  
5 believed to increase the transcription efficiency. Preferably, a nucleotide sequence coding for signal peptide for extracellular secretion is added just upstream of the nucleotide sequence coding for an interferon conjugate, extracellularly to secrete the  
10 interferon conjugate, which can be then recovered in a supernatant of culture broth.

To prepare a large amount of the above-mentioned DNA construct so that the DNA construct can be easily introduced into animal cells, the DNA construct  
15 preferably includes an E. coli replication origin and a drug resistance gene. The replication origin is preferably, but not limited to, that derived from a colicin E1 plasmid, for example, pBR22, or a related plasmid. The drug resistance gene is, for example, an  
20 ampicillin resistant gene, tetracycline resistant gene, or kanamycin resistant gene. Moreover, the DNA construct can contain a replication origin responsible for autonomous replication in host cells, for example, a replication origin of SV40 virus or a replication origin  
25 of polyoma virus. These various DNA fragments are linked to construct an interferon conjugate expression vector.

The construction of a vector DNA can be carried out according to a conventional procedure (T. Maniatis et  
30 al., "Molecular Cloning", p86-96, 1982).

Animal cells into which a vector DNA will be introduced may be cells derived from humans, monkeys, Chinese hamsters, mice, etc., although since a human interferon conjugate is the target compound, human cells  
35 are preferably used. Human cells wherein growth is not inhibited by a peptide having glycoside chains produced are used. The human cells are preferably cells derived

from human lung cancer, especially PC8 and PC12 cell lines (M. Kinjo et al., Br. J. Cancer, 39, 15, 1979).

Vector DNA can be introduced into host cells according to a known calcium phosphate method (F.L. 5 Graham et al., Virology, 54, 536, 1973).

Cells into which a plasmid for expression of an interferon conjugate has been introduced can be efficiently obtained by cotransformation of host cells with the plasmid and G418 resistance gene expression 10 vector pSV2neo (P.J. Southern et al., J. Mol. Appl. Genet., 1, 327, 1982) or pNE05' (M. Lusky et al., Cell, 36, 391, 1984), because the cotransformed cells can survive in a medium containing G418, in which non-cotransformed cells cannot survive.

15 The thus-obtained transformant is cultured in, for example, a medium containing fetal calf serum, extracellularly to produce an interferon conjugate, which is then recovered in a pure form from a supernatant of the cultured broth according to the same procedure as 20 described for recovery from the E. coli extract. The thus-obtained interferon conjugate is a polypeptide having glycoside chains.

Since the interferon conjugate prepared as described above binds to an anti-human interferon- $\beta$  antibody and 25 to an anti-human interferon- $\gamma$  antibody, it has the antigenecities of both interferons- $\beta$  and - $\gamma$ . Moreover, a neutralization test thereof with anti-human interferon- $\beta$  antibody and anti-human interferon- $\gamma$  antibody showed that a single polypeptide exhibits both 30 interferon- $\beta$  activity and interferon- $\gamma$  activity.

As described above, the present invention provides a new type of interferon conjugate comprising a region exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological activities of interferon- $\gamma$ , 35 which is produced by recombinant cells containing DNA coding for interferon- $\beta$  and interferon- $\gamma$ .

The interferon conjugate of the present invention

is a single polypeptide carrying biological actions which were originally carried separately on interferon- $\beta$  and interferon- $\gamma$ . Therefore, the present interferon conjugate has a broader action spectra, such as an antiviral spectrum and an anti-cell proliferation spectrum broader than those of conventional interferon. These properties make the present interferon conjugate valuable as an antiviral pharmaceutical or an antitumor pharmaceutical.

Moreover, since the present interferon conjugate exhibits the synergism conventionally provided by a mixture of interferon- $\beta$  and interferon- $\gamma$ , it provides remarkably enhanced activity per molecule in comparison to conventional interferons. Although such synergism can be obtained in vitro by only mixing interferon- $\beta$  and - $\gamma$ , since interferons- $\beta$  and - $\gamma$ , would show different in vivo pharmacodynamics, the coexistence of both interferons at a target site is not assured in vivo. This means that the mixture of both interferons- $\beta$  and - $\gamma$  would not exhibit the synergism. On the contrary, in the present interferon conjugate, since a single molecule provides the synergism, the problem of different pharmacodynamics does not exist. This also makes the present interferon conjugate valuable as an antiviral pharmaceutical and an antitumor pharmaceutical.

Finally, at present, to obtain interferon- $\beta$  activity and interferon- $\gamma$  activity, separate processes for the production of interferons- $\beta$  and - $\gamma$  must be carried out. On the contrary, according to the present invention, one production process provides an interferon conjugate carrying both interferon- $\beta$  activity and interferon- $\gamma$  activity. This gives an economical advantage to the present invention. The present interferon conjugate can be used as such, or can be easily cleaved into interferon- $\beta$  and interferon- $\gamma$ , which then can be used separately or as a mixture. In all cases the above-mentioned advantage still remains.

Examples

The present invention will now be further illustrated by, but is by no means limited to, the following examples.

- 5 Common procedures for gene manipulation used in the following Reference example and Examples are those described in Maniatis et al., "Molecular Cloning", Cold Spring Harbor Laboratory, 1982.

10 Prior to illustrating Examples of the present invention, the construction processes of a plasmid for the expression of human interferon- $\beta$  and a plasmid for the expression of human interferon- $\gamma$  are described as a Reference example. These plasmids are used as starting plasmids for the construction of the present plasmids.

15 Reference example

- (1) Construction of plasmid pKM6 for expression of human interferon- $\beta$ .

A plasmid pTuIFN $\beta$ -5 for the expression of human interferon- $\beta$  constructed according to a reported  
20 procedure (Taniguchi, Seikagaku, 54, 363-377, 1982) was digested with HindIII, and the digestion product was treated with a T4 DNA polymerase Klenow fragment to make blunt ends, linked to a BglII linker, digested with BglII, and self-circularized using a T4 DNA ligase to  
25 obtain a plasmid pYO-10. The plasmid pYO-10 was digested with SalI and ClaI, and the digestion product was subjected to agarose gel electrophoresis to isolate a DNA fragment of about 830 bp. This DNA fragment was inserted into a ClaI-SalI region of the plasmid p6huy-A2  
30 described in Japanese Unexamined Patent Publication No. 61-19488 to construct a plasmid pKM6 shown in Fig. 3.

- (2) Construction of plasmid p6huy-N1 for expression of human interferon  $\gamma$ .

35 Lymphocyte derived from human tonsils was treated with phytohemagglutinin (PHA) and 12-o-tetradecanoyl-phorbol-13-acetate (TPA) to induce the production of human interferon- $\gamma$  (Vilvek et al.,



Infection and Immunity, 34, 131, 1983), and then mRNA was prepared from the treated cells. The preparations of mRNA and cDNA, and the cloning of cDNA in a plasmid were carried out according to known procedures (Okayama  
5 et al., Molecular and Cellular Biology, 3, 280, 1983). The DNA library thus obtained was subjected to colony hybridization using a synthetic oligonucleotide probe having the sequence: 5'-AGGACAACCATTACT-3' corresponding to a region near the 3'-terminal of a known human  
10 interferon- $\gamma$  structural gene (Goeddel et al., Nature, 295, 503-509, 1982) to obtain a plasmid pIFN- $\gamma$ 15 containing a cDNA coding for human interferon- $\gamma$ . Next, after the plasmid pIFN- $\gamma$ 15 was digested with NdeI, and BamHI, a DNA fragment of about 0.9 kb was isolated by  
15 agarose gel electrophoresis.

On the other hand, two oligonucleotides 5'-CGATGCAGGACCCA-3' and 5'-TATGGGTCCTGCAT-3' were synthesized, and after phosphorylation at the 5'-end with a T4 polynucleotide kinase, the oligonucleotides  
20 were mixed to make a concentration of about 8 pmol/ $\mu$ l for each oligonucleotide. The mixture was then heated at 65°C for 3 minutes, cooled rapidly, heated again at 65°C for 3 minutes, and then allowed to stand at a room temperature to be cooled gradually and to anneal the  
25 oligonucleotides.

7 pmoles of this oligonucleotide, 0.3 pmole of NdeI-BamHI fragment from plasmid pIFN $\gamma$ -15, and 0.1 pmole of a DNA fragment of about 4200 bp prepared by digestion of the plasmid pKM6 described in paragraph (1) with ClaI  
30 and BamHI followed by isolation by agarose gel electrophoresis were mixed, and after ligation the ligation mixture was used to transform E. coli MC1061 (Casadaban et al., J. Mol. Biol., 138, 179-207, 1980). The transformants selected for ampicillin resistance were sub-  
35 jected to colony hybridization using an oligonucleotide probe 5'-TATGGGTCCTGCAT-3', to obtain a plasmid p6hur-N1 for expression of human interferon- $\gamma$  (Fig. 4).

Next, to link the interferon- $\beta$  structural gene and interferon- $\gamma$  structural gene, restriction enzyme sites are introduced to the 5'-end or 3'-end of each structural gene.

5           (3) Construction of plasmid pKM6-Cxho

The structure of the plasmid pKM6-Cxho is shown in Fig. 5. The plasmid pKM6 constructed as described in paragraph (1) was digested with BstEII and BamHI, and an adapter oligonucleotide having the following sequence:

GTTACCTCCGAAACTCGAGCTGA

GAGGCTTTGAGCTCGACTCTAG

was linked to the cleaved plasmid to construct the plasmid pKM6-cxho. The adapter was prepared according to the same procedure as described in paragraph (2). When the plasmid pKM6-Cxho is digested with XhoI and the protruding nucleotides are eliminated, a codon AAC coding for aspartic acid present at the C-terminal of human interferon- $\beta$  is exposed.

20           (4) Construction of plasmid p6huyN1-CKpn

The structure of the plasmid p6huyN1-CKpn is shown in Fig. 6. The plasmid p6huy-N1 constructed as described in paragraph (2) was digested with ClaI and BamHI, and after agarose gel electrophoresis, a DNA fragment of about 4200 bp and a DNA fragment of about 1050 bp were isolated. The DNA fragment of about 1050 bp was further digested with HinfI, and after agarose gel electrophoresis a DNA fragment of about 400 bp was isolated. The ClaI-BamHI fragment of about 4200 bp prepared as described above, the ClaI-HinfI fragment of about 400 bp, and an adapter of the following sequence:

(5')-AGTCAGATGCTGTTTGGGGTGGAGGTGCATCCAGGTACCATGAGATCTG

GCTACGACAAAGCGCCAGCTGCAAGTAGGGTCCATGGTACTCTAGACTAG-(5')

35 were mixed and ligated, and the ligation mixture was used to transform E. coli MC1061. The above-mentioned adapter was prepared from six oligonucleotides according

t the same procedure as described in paragraph (2).  
The transformants selected for ampicillin resistance  
were subjected to colony hybridization using an  
oligonucleotide probe 5'-GATCCAGATCTCATG-3' to obtain  
5 four positive clones from 118 clones. The positive  
clones contained plasmid p6hu $\gamma$ -CKpn. When the thus-  
prepared plasmid p6hu $\gamma$ -CKpn is digested with KpnI, and  
protruding nucleotides are eliminated, a codon CAG  
coding for glutamine present at the C-terminal of human  
10 interferon- $\gamma$  is exposed.

(5) Construction of plasmid p6hu $\gamma$ N1 $\Delta$ BS-NH $\alpha$ n

The structure of plasmid p6hu $\gamma$ N1 $\Delta$ BS-NH $\alpha$ n is  
shown in Fig. 7. The plasmid p6hu $\gamma$ -N1 constructed as  
described in paragraph (2) was digested with BstEII, the  
15 resulting cohesive ends were changed to blunt ends using  
a DNA polymerase I Klenow fragment, and the resulting DNA  
fragment was linked to a SalI linker. After digestion  
with SalI, the DNA fragment was self-circularized using  
the T4 DNA ligase to obtain p6hu $\gamma$ N1- $\Delta$ BS. The plasmid  
20 p6hu $\gamma$ N1- $\Delta$ BS was digested with NdeI and SalI, and after  
agarose gel electrophoresis, a DNA fragment of about  
3700 bp was isolated.

On the other hand, the plasmid pKM6 constructed  
as described in paragraph (2) was digested with EcoRI  
25 and SalI, and after agarose gel electrophoresis, a DNA  
fragment of about 3700 bp was isolated.

Two DNA fragments prepared as described above  
were linked with an adapter oligonucleotide of the  
following sequence:

AATTGCGCAGGACCCA

CGCGTCCTGGGTAT

to obtain plasmid p6hu $\gamma$ N1 $\Delta$ BS-NH $\alpha$ n. When the plasmid  
p6hu $\gamma$ N1 $\Delta$ BS-NH $\alpha$ n is digested with HinPI, and protruding  
nucleotides are eliminated, a codon CAG coding for  
35 glutamine present at the N-terminal of human interferon- $\gamma$   
is exposed.

Example 1. Construction of plasmid ptrp6huIFN- $\gamma$ 8

for expression of interferon- $\gamma$ - $\beta$   
conjugate

A process for the construction of a plasmid ptrp6huIFN- $\gamma$  $\beta$  is shown in Fig. 8, wherein 30  $\mu$ g of  
5 plasmid pKM6 was digested with ClaI, and the digestion  
product was treated with 15 units of mung bean nuclease  
to change cohesive ends to blunt ends. The DNA fragment  
thus-obtained was further digested with BglII, and after  
10 agarose gel electrophoresis, a DNA fragment of about  
500 bp was isolated. On the other hand, plasmid  
p6hu $\gamma$ N1-CKpn was digested with KpnI, the cohesive ends  
were changed to blunt ends using T4 DNA polymerase, and  
after digestion with BamHI followed by agarose gel  
15 electrophoresis, a DNA fragment of about 4800 bp was  
obtained. The DNA fragments thus-prepared were mixed  
and ligated using T4 DNA ligase, and the reaction  
mixture was used to transform E. coli HB101 (Boyer et  
al., J. Mol. Biol., 41, 459-472, 1969). The resulting  
ampicillin resistant transformants were subjected to  
20 colony hybridization using as a probe a  $^{32}$ P-labeled DNA  
fragment prepared by nick-translating a ClaI-BglII  
fragment of plasmid pKM6, to obtain 6 positive clones  
among 56 clones. Plasmid DNA was extracted from each of  
these clones, and a restriction enzyme cleavage map was  
25 made. As a result, it was confirmed that plasmids from  
all positive clones had a structure as shown in Fig. 8.  
Moreover, plasmid DNA derived from a representative  
clone  $\gamma$ 86 was digested with SalI, and resulting DNA  
fragments were inserted to M13 phage to determine a  
30 nucleotide sequence of the plasmid DNA. As a result, in  
this plasmid, an IFN- $\gamma$  structural gene was linked with  
an IFN- $\beta$  structural gene in frame. This desired plasmid  
was designated as plasmid ptrp6huIFN- $\gamma$  $\beta$ , and a trans-  
formant E. coli was designated as E. coli HB101  
35 (ptrp6huIFN- $\gamma$  $\beta$ ).

Example 2. Construction of plasmid ptrp6huIFN- $\beta$ a  
for expression of interferon- $\beta$ -a

conjugate

A process for the construction of a plasmid ptrp6huIFN- $\beta$  is shown in Fig. 9, wherein 20  $\mu$ g of plasmid pKM6-Cxho was digested with Xho I, and the digestion product was treated with 15 units of mung bean nuclease at 37°C for 15 minutes to make blunt ends, and then digested with SalI. The reaction mixture was subjected to agarose gel electrophoresis to isolate a DNA fragment of about 4500 bp. On the other hand, 30  $\mu$ g of p6hu $\gamma$ Nl $\Delta$ BS-NHln was digested with HinfI, the digestion product was treated with 30 units of mung bean nuclease at 37°C for 15 minutes, and digested with SalI, and after agarose gel electrophoresis a DNA fragment of about 860 bp was isolated.

The DNA fragments thus-prepared were mixed and ligated using the T4 DNA ligase, and the reaction mixture was used to transform E. coli HB101. Among the resulting ampicillin resistant transformants, 50 clones were subjected to colony hybridization using a oligonucleotide probe 5'-AGTCAGATGCTGTTTC-3', which is the same oligonucleotide used for the construction of plasmid p6hu $\gamma$ Nl-CKpn, to obtain 28 positive clones. A plasmid DNA was isolated from a representative clone  $\beta$ Y31, and a restriction enzyme cleavage map was made. As a result, a map as shown in Fig. 9 was obtained. Moreover, a BstEII-SalI fragment from the plasmid isolated from the clone  $\beta$ Y31 was cloned in phage M13 to determine the nucleotide sequence. As a result, in the plasmid, an IFN- $\beta$  structural gene was linked with an IFN- $\gamma$  structural gene in frame. This plasmid was designated as plasmid ptrp6huIFN- $\beta$  $\gamma$ , and E. coli transformed with the plasmid was designated as E. coli HB101 (ptrp6huIFN- $\beta$  $\gamma$ ).

Example 3. Construction of plasmid ptrphuIFN- $\gamma$ c $\beta$  for expression of interferon- $\gamma$ c $\beta$  conjugate

A process for the construction of a ptrphuIFN- $\gamma$ c $\beta$

is shown in Fig. 10. A plasmid pKM6 was digested with  
 ClaI and then with BglII, and after agaros gel  
 electrophoresis a DNA fragment of about 500 bp was  
 isolated. On the other hand, a DNA fragment coding for  
 5 a spacer peptide was prepared from four synthetic  
 oligonucleotides according to the same procedure as  
 described in paragraph (2) of the Reference Example. 10  
 pmoles of this DNA fragment was mixed with the ClaI-BglII  
 fragment of pKM6 isolated as described above and the DNA  
 10 fragment of about 4800 bp derived from plasmid  
 p6huγN1-CKpn as described in Example 1, these DNA  
 fragments were ligated using the T4 DNA ligase, and the  
 reaction mixture was used to transform E. coli HB101.  
 82 clones of the resulting ampicillin resistant trans-  
 15 formants were subjected to colony hybridization using  
 the probe described in Example 1 to obtain three positive  
 clones. Plasmid DNAs were isolated from the three  
 clones, and restriction enzyme cleavage maps were made.  
 As a result, only one clone contained a plasmid having a  
 20 desired structure. This plasmid was designated as  
 plasmid ptrp6huIFN-γc8, and E. coli transformed with  
 this plasmid was designated as E. coli HB101  
 (ptrp6huIFN-γc8).

25        Example 4.    Culturing and production of interferon  
    conjugate

Transformants obtained in Examples 1 to 3 were  
 inoculated to LB medium (1.0% bactotrypton, 0.5% yeast  
 extract, 0.5% sodium chloride, 0.1% glucose, pH 7.2 with  
 sodium hydroxide) supplemented with 100 μg/ml tryptophan,  
 30 and 100 μg/ml ampicillin; and cultivation was carried  
 out at 30°C for 8 hours. This culture was then  
 inoculated to M9 medium (0.3% mono potassium phosphate,  
 0.6% disodium phosphate, 0.1% ammonium chloride, and  
 0.5% sodium chloride, and further, separately sterilized  
 35 1 μg/ml Vitamine B<sub>1</sub> and 0.1 mM magnesium sulfate)  
 supplemented with 1.0% glucose and 1.0% casamino acids,  
 in an amount of 10% by volume relating to a volume of

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the M9 medium, and cultivation was continued. After about 10 hours, indoleacrylic acid was added to the final concentration of 10 µg/ml, and cultivation was carried out for an additional 8 hours. During the cultivation, if necessary, 40% glucose aqueous solution was added to the culture medium to prevent depletion of the glucose. Moreover, during the cultivation, a 14% aqueous ammonia solution was added to the culture medium to maintain the pH value at 6.0 to 7.0.

10 After the cultivation, 2 ml of the cultured broth was centrifuged at 10,000X g for 4 minutes to collect cells. The cells were washed with physiological saline, and the washed cells were suspended in 1 ml of 50 mM Tris-HCl buffer (pH 7.5) containing 3 mg of lysozyme,  
15 2 mM EDTA, 30 mM sodium chloride, and 20% glycerol, and the suspension was allowed to stand in ice for 60 minutes. The freezing and thawing of the suspension were repeated three times to disrupt the cells, and the disrupted mixture was centrifuged at 30,000X g for  
20 20 minutes to eliminate cell debris. A supernatant thus obtained was used as a test sample for measuring.

The antiviral activity of interferon was measured according to "Interferon no Kagaku" (Science of Interferon) edited by Shigeyasu Kobayashi, pl3-20, Kodan  
25 Sha, 1985, by the CPE<sub>50</sub> inhibition method using the FL cells-sindbis virus system. As a standard for measurement of the activity, an IFN-γ preparation was used which was produced by recombinant cells and calibrated using NIH natural IFN-γ Gg23-901-530. For  
30 reference, interferon crude extracts prepared according to the above-mentioned procedure from E. coli HB101 containing plasmid pKM6, which expresses human interferon-β, and from E. coli HB101 containing plasmid p6hur-N1, which express human interferon-γ, were tested  
35 for their antiviral activity. The results are shown in Table 1.

Table 1

Strain	Antiviral Activity per Extract (U/ml)
<u>E. coli</u> HB101 (ptrp6huIFN- $\gamma$ 8)	$3.9 \times 10^4$
<u>E. coli</u> HB101 (ptrp6huIFN- $\beta$ $\gamma$ )	$1.6 \times 10^4$
<u>E. coli</u> HB101 (ptrp6huIFN- $\gamma$ c8)	$7.7 \times 10^4$
<u>E. coli</u> HB101 (pKM6)	$3.1 \times 10^5$
<u>E. coli</u> HB101 (p6hu $\gamma$ -N1)	$4.1 \times 10^4$

As seen from Table 1, each preparation exhibited an  
20 antiviral activity characteristic for the expected  
interferon.

Example 5. Measurement of Molecular Weight

1 ml of cultured broth prepared according to the  
same procedure as described in Example 4 was centrifuged  
25 at 10,000X g for 4 minutes to collect cells. The cells  
were suspended in 500  $\mu$ l of 62.5 mM Tris-HCl buffer  
(pH 6.8) containing 5% 2-mercaptoethanol and 2% sodium  
dodecylsulfate, and the suspension was heated in a  
boiling water bath for 5 minutes and allowed to cool.  
30 Then, to the heat-treated suspension was added 50  $\mu$ l of  
62.5 mM Tris-HCl buffer (pH 6.8) containing 0.05% bromo  
phenol blue and 70% glycerol, to prepare a sample for  
electrophoresis. SDS-polyacrylamide gel electrophoresis  
was carried out according to Laemmli's method (Nature,  
35 227, 680, 1970). The gel concentration was 15%. As  
maker proteins, lysozyme having a molecular weight of  
14,400, trypsin inhibitor having a molecular weight of



21,500, carbonic anhydrase having a molecular weight of 31,000, ovalbumin having a molecular weight of 45,000, bovine serum albumin having a molecular weight of 66,200, and phosphorylase B having a molecular weight of 92,500 were used. After electrophoresis, the gel was stained with Coomassie Brilliant Blue R250 to detect proteins. Another gel obtained by simultaneous electrophoresis was used to transport proteins on the gel to a nitrocellulose sheet. The nitrocellulose sheet was reacted with commercially available anti-human interferon- $\beta$  immunoglobulin or anti-human interferon- $\gamma$  immunoglobulin, and then reacted with peroxidase-labeled protein A to determine the position of the interferon conjugate. By comparing a result of the above-mentioned Western blotting with the positions of electrophoretically migrated marker proteins, it was found that IFN- $\gamma\beta$  and IFN- $\beta\gamma$  have a molecular weight of about 37,000, and IFN- $\gamma\beta$  has a molecular weight of about 38,000. That is, it was confirmed that interferon conjugates of the present invention substantially consist of human interferon- $\beta$  having a molecular weight of about 20,000 and human interferon- $\gamma$  having a molecular weight of about 17,000. Note, about 1000 of a molecular weight in IFN- $\gamma\beta$  is assigned to a spacer peptide contained in the IFN- $\gamma\beta$ .

Example 6. Neutralization of product with antibody

A crude interferon extract from the E. coli HB101 (ptrp6huIFN- $\gamma\beta$ ) prepared according to the procedure described in Example 4 was diluted five-fold with an Eagle's minimum essential medium containing 5% calf serum and 10 mM Hepes (pH 7.3). To 1 ml of the interferon solution thus-prepared, 1 ml of anti-IFN- $\beta$  rabbit antiserum diluted 50-fold with the same medium (neutralization value 2700 U/ml) or 1 ml of anti-IFN- $\gamma$  rabbit antiserum diluted 50-fold with the same medium (neutralization value 2000 U/ml) was added, and after the mixture was incubated at 37°C for 30 minutes, the

antiviral activity was measured. For control, a reaction mixture wherein the antiserum was replaced by the medium, and a reaction mixture wherein 0.5 ml of the anti-IFN- $\beta$  rabbit antiserum solution and 0.5 ml of the anti-IFN- $\gamma$  rabbit antiserum solution were added were prepared, and the antiviral activity was measured in same manner. The results are set forth in Table 2.

Table 2

Antiserum	Antiviral Activity (U/ml)
Control (no antiserum)	$6.0 \times 10^3$
Anti-IFN- $\beta$ antiserum	$1.3 \times 10^3$
Anti-IFN- $\gamma$ antiserum	$1.7 \times 10^3$
Anti-IFN- $\beta$ antiserum + and -IFN- $\gamma$ antiserum	$8.1 \times 10$

As seen from Table 2, each of the anti-IFN- $\beta$  antiserum and the anti-IFN- $\gamma$  antiserum partially neutralized antiviral activity, and a combination of anti-IFN- $\beta$  antiserum and anti-IFN- $\gamma$  antiserum substantially completely neutralized all antiviral activity. This means that the present interferon- $\gamma\beta$  conjugate has both an interferon- $\beta$  action and an interferon- $\gamma$  action. Moreover, although  $2.7 \times 10^3$  U of anti-IFN- $\beta$  antiserum neutralized the IFN- $\beta$  activity with  $1.3 \times 10^3$  U of IFN- $\gamma$  activity remaining and  $2.0 \times 10^3$  U of anti-IFN- $\gamma$  antiserum neutralized the IFN- $\gamma$  activity with  $1.7 \times 10^3$  U of IFN- $\beta$  activity remaining, the not-neutralized control showed  $6.0 \times 10^3$  U activity. That is, antiviral activity of the control ( $6.0 \times 10^3$ ) is higher than total amount of remaining activities ( $1.3 \times 10^3$  U +  $1.7 \times 10^3$  U). This means that the present interferon- $\gamma\beta$  conjugate exhibits synergism of IFN- $\beta$

activity and IFN- $\gamma$  activity.

Example 7. Construction of plasmid ptrp6huIFN- $\beta$ cy for expression of interferon  $\beta$ cy

A process for the construction of a plasmid ptrp6huIFN- $\beta$ cy is shown in Fig. 12, wherein 20  $\mu$ g of plasmid pKM6-Cxho was digested with XhoI, the digestion product was treated with 15 units of mung bean nuclease at 37°C for 15 minutes to make blunt ends, and then digested with SalI. The reaction mixture was subjected to agarose gel electrophoresis to isolate a DNA fragment of about 4500 bp. On the other hand, 30  $\mu$ g of plasmid p6hu $\gamma$ N1 $\Delta$ BS-NHin was digested with HinPI and SalI, and after agarose gel electrophoresis, a DNA fragment of about 860 bp was isolated. The above-mentioned two DNA fragments were mixed with 10 pmoles of DNA fragment coding for a spacer peptide obtained according to the procedure as described in Example 3, these three DNA fragments were ligated with T4 DNA ligase, and the ligation mixture was used to transform E. coli HB101. The 204 ampicillin resistant transformants thus obtained were subjected to colony hybridization using as a probe that shown in Example 2 or an oligonucleotide 5'-CGTTACCGACTTAGCA-3' used for preparation of the DNA fragment coding for the spacer peptide. As a result, two clones were found to be positive, and plasmids from these clones were analyzed by restriction enzymes. One clone contained a desired plasmid, and this plasmid was designated as plasmid ptrp6huIFN- $\beta$ cy, and E. coli transformed with this plasmid was designated as E. coli HB101 (ptrp6huIFN- $\beta$ cy). According to the procedures described in Example 4, the E. coli transformant was cultured and an extract prepared, and then the antiviral activity was measured. An antiviral activity of  $3.9 \times 10^4$  U/ml extract was found.

Example 8. Neutralization of product with antibody

According to the same procedure as described in Example 6, a crude interferon extract from E. coli HB101

(ptrp6huIFN- $\gamma$ c $\beta$ ) was neutralized with antisera. For comparison, an interferon mixture consisting of 8600 U/ml of IFN- $\beta$  and 2400 U/ml of IFN- $\gamma$ , prepared by mixing an IFN- $\beta$  and an IFN- $\gamma$  in approximately the same molecular numbers, were tested in the same manner. The results are shown in Table 3.

Table 3

IFN	Antiserum		Antiviral activity (U/ml)
	Anti-IFN- $\beta$	Anti-IFN- $\gamma$	
IFN mixture	-	-	19000
	o	-	930
	-	o	12000
	o	o	<27
IFN- $\gamma$ c $\beta$	-	-	22000
	o	-	2400
	-	o	11000
	o	o	61

As can be seen from Table 3, in the interferon- $\gamma$ c $\beta$  conjugate (IFN- $\gamma$ c $\beta$ ), each of the anti-IFN- $\beta$  antiserum and anti-IFN- $\gamma$  antiserum partially neutralized the antiviral activity thereof, and a combination of both antisera substantially completely neutralized all antiviral activity. This means that correctly folded IFN- $\beta$  and IFN- $\gamma$  molecules are liked to yield both the IFN- $\beta$  activity and IFN- $\gamma$  activity are exhibited by a single polypeptide.

Moreover, from the comparison of the activities resulting from neutralization with the anti-IFN- $\beta$  or anti-IFN- $\gamma$  with the activity resulting from

neutr lization with f a combination of anti-IFN- $\beta$  and anti-IFN- $\gamma$ , it appears that the present IFN- $\gamma$ c $\beta$  conjugat als exhibits the synergism of the IFN- $\beta$  activity and the IFN- $\gamma$  activity.

5        Example 9.

      A.    Construction of vector for expression of human interferon- $\beta$

      pSV $\beta$  is a vector derived from the human interferon- $\gamma$  expression vector pSV2IFN $\beta$  (Japanese  
10 Unexamined Patent Publication No. 61-52283) by deleting a sequence inhibiting replication of the vector in a eukaryotic cell (M. Losky et al., Nature, 293 77, 1981). The vector pSV $\beta$  was constructed as follows.

      First, a PvuII site positioned in the pSV2IFN $\beta$   
15 upstream from the SV40 early promoter was replaced by a SalI site using a SalI linker, and the modified vector was cleaved with SalI and BamHI to isolate a 1.7 Kb DNA fragment required for expression of the human interferon- $\beta$ .

20        Next, vector pML2d (M. Lusky et al., Nature, 293, 79, 1981) derived from pBR322 by deleting a sequence inhibiting replication of the vector in the eukaryotic cell was cleaved with SalI and BamHI to isolate a long DNA fragment.

25        These two DNA fragments were ligated using T4 DNA ligase to obtain pSV $\beta$ .

      B.    Construction of vector pMTV $\beta$  for expression of human interferon- $\beta$

      The vector pSV $\beta$  constructed in Section A was  
30 cleaved with SalI, and the SalI site in the vector was replaced by a HindIII site using a HindIII linker, and the modified vector was cleaved with HindIII to isolate a 3.8 Kb DNA fragment which does not contain the SV40 early promoter. The DNA fragment thus obtained was then  
35 treated with E. coli alkaline phosphatase to eliminate a terminal phosphate group.

      Next, vector pMTVdhfr (F. Lee et al., Nature,

294, 228, 1982) containing MMTV promoter was cleaved with a restriction enzyme HindIII to isolate a 1.4 Kb DNA fragment containing MMTV promoter.

These two DNA fragments were ligated using T4 DNA ligase to obtain pMTV $\beta$ .

C. Construction of vector pMTV $\gamma$  for expression of human interferon- $\gamma$

pMTV $\gamma$  is a vector wherein a human interferon- $\gamma$  gene is positioned under control by an MMTV promoter.

The vector pMTV $\beta$  constructed in Section B was cleaved at a HindIII site downstream from the MMTV promoter and at the BglII site downstream from the human interferon gene, and after the resulting fragments were blunt-ended using a DNA polymerase I Klenow fragment, a 3.9 Kb DNA fragment containing the MMTV promoter was isolated.

This DNA fragment was ligated with a 0.8 Kb DNA fragment containing a human interferon- $\gamma$  gene, which was obtained from pSVIFN $\gamma$  (Japanese Unexamined Patent Publication No. 61-52286) by cleaving with DpnI, using a T4 DNA ligase to obtain pMTV $\gamma$ .

D. Construction of vector pMTV(SV) $\gamma$  for expression of human interferon- $\gamma$

pMTV(SV) $\gamma$  is a vector derived from pMTV $\gamma$  by inserting an SV40 early promoter upstream of the MMTV promoter. The vector pMTV(SV) $\gamma$  was constructed as follows.

pMTV $\gamma$  constructed in Section C was cleaved with SalI, and after the resulting DNA fragment was blunt-ended using a DNA polymerase I Klenow fragment, a terminal phosphate group was eliminated by BAP treatment.

Next, pSV2IFN $\beta$  (Japanese Unexamined Patent Publication 61-52283) was cleaved with PvuII and HindIII to isolate a 0.3 Kb DNA fragment containing the SV40 early promoter, and the fragment was blunt-ended by treating with a DNA polymerase I Klenow fragment.

These two DNA fragments were ligated using T4

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DNA ligase to obtain pMTV(SV) $\gamma$ .

E. Construction of plasmid pMTV(SV) $\gamma$  $\cdot$  $\beta$  for  
expression of human interferon- $\gamma$  $\cdot$  $\beta$  conjugate  
in animal cells (Fig. 13)

5 pMTV(SV) $\gamma$  $\cdot$  $\beta$  is a vector wherein the human  
interferon- $\gamma$  gene in pMTV(SV) $\gamma$  has been replaced by the  
human interferon- $\gamma$  $\cdot$  $\beta$  conjugate gene, and was constructed  
as follows.

10 10  $\mu$ g of ptrp6huIFN- $\gamma$  $\beta$  constructed according  
to the procedure as described in Example 1 was digested  
with NdeI and DpnI, and the digestion mixture was  
subjected to agarose gel electrophoresis to isolate a  
DNA fragment of about 1300 bp.

15 On the other hand, pMTV(SV) $\gamma$  constructed in  
Section D was digested with BglII, and the resulting  
fragment was blunt-ended by treating with DNA  
polymerase I Klenow fragment, followed by digestion with  
NdeI, and the reaction mixture was subjected to agarose  
gel electrophoresis to isolate a DNA fragment of about  
20 5100 bp.

These DNA fragments were mixed and ligated  
with T4 DNA ligase to obtain pMTV(SV) $\gamma$  $\cdot$  $\beta$ .

Example 10. Construction of plasmid pMTV(SV) $\gamma$  $\cdot$  $\beta$   
for expression of human interferon- $\gamma$  $\cdot$  $\beta$   
conjugate in animal cells

25 Plasmid pMTV(SV) $\gamma$  $\cdot$  $\beta$  is a vector wherein a human  
interferon- $\gamma$  gene in pMTV(SV) $\gamma$  has been replaced by a  
human interferon- $\gamma$  $\cdot$  $\beta$  conjugate gene, and was constructed  
as follows.

30 10  $\mu$ g of ptrp6huIFN- $\gamma$  $\cdot$  $\beta$  constructed according to  
the procedure as described in Example 3 was digested  
with NdeI and DpnI, and the reaction mixture was  
subjected to agarose gel electrophoresis to isolate a  
DNA fragment of about 1300 bp. On the other hand,  
35 pMTV(SV) $\gamma$  constructed in Section D was digested with  
BglII, and the resulting DNA fragment was blunt-ended by  
treating with a DNA polymerase I Klenow fragment,

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followed by digestion with NdeI, and the reaction mixture was subjected to agarose gel electrophoresis to isolate a DNA fragment of about 5100 bp.

These DNA fragments were mixed, and ligated using T4 DNA ligase to obtain pMTV(SV) $\gamma$ c $\beta$ .

Example 11. Transformation of PC12 cell with  
MTV(SV) $\gamma$ c $\beta$

4  $\mu$ g of vector pMTV(SV) $\gamma$ c $\beta$  constructed in Example 9 and 0.4  $\mu$ g of GC18 resistance gene expression vector pSV2neo (J. Southern et al., J. Mo. Appl. Genet., 1, 327, 1982) were introduced into about  $10^6$  cells of PC12 cell line (M. Kinjo et al., Br. J. Cancer, 39, 15, 1979) derived from human lung cancer, according to the calcium phosphate method. The cells thus-prepared were cultured in a selective medium (RPMI 1640 medium supplemented with 10% fetal calf serum and 100  $\mu$ g/ml kanamycine; Nissui Seiyaku, Japan) containing 400  $\mu$ g/ml of a protein synthesis inhibitor, GC18 (GIBCO) to obtain 24 transformant clones. The antiviral activity of the cultured supernatants was tested using FL cells-sindbis virus system according to the CPE<sub>50</sub> inhibition method described in Example 4. As a result, supernatants from 22 clones exhibited the antiviral activity, as shown in Table 4.



Table 4

pMTV(SV)γ-8/PC12	
Clone	Antiviral activity (U/ml)
1	18500
2	1100
3	600
4	1500
5	< 80
6	1300
7	200
8	2300
9	200
10	< 80
11	1000
12	2500
13	900
14	1400
15	500
16	400
17	< 80
18	< 80
19	300
20	800
21	200
22	300
23	200
24	600

Example 12. Transformati n of PC12 cells with  
pMTV(SV) $\gamma$ c8

4  $\mu$ g of pMTV(SV) $\gamma$ c8 constructed in Example 10 and  
0.4  $\mu$ g of pSV2neo (see Example 11) were introduced into  
5 about  $10^6$  cells of the PC12 cell line by a calcium  
phosphate method according to the same procedure as  
described in Example 11. The cells thus-prepared were  
cultured in a selective medium (RPMI 1640 medium  
supplemented with 10% fetal calf serum and 100  $\mu$ g/ml  
10 kanamycine; Nissui Seiyaku, Japan) to obtain 26  
transformed clones. The antiviral activity of the  
cultured supernatants was tested by the CPE<sub>50</sub> inhibition  
method using the FL cells-sindbis virus system according  
to the same procedure as described in Example 11. As a  
15 result, supernatants from 20 clones exhibited the  
antiviral activity as shown in Table 5.

Table 5

pMTV(SV) $\gamma$ CB/PC12	
Clone	Antiviral activity (U/ml)
1	400
2	< 60
3	< 60
4	3800
5	1200
6	< 60
7	6400
8	200
9	< 80
10	400
11	200
12	900
13	500
14	700
15	< 80
16	1300
17	21450
18	130
19	5300
20	1600
21	200
22	< 80
23	1200
24	8600
25	300
26	500

CLAIMS

1. An interferon conjugate comprising in a single molecule a region exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological activities of interferon- $\gamma$ .
- 5 2. An interferon conjugate according to claim 1, which is a biologically active polypeptide having no glycoside chain.
3. An interferon conjugate according to claim 1, which is a biologically active polypeptide having  
10 glycoside chains.
4. An interferon conjugate according to any one of claims 1 to 3, wherein the region exhibiting biological activities of interferon- $\beta$  and the region exhibiting biological activities of interferon- $\gamma$  have  
15 been linked via a spacer.
5. An interferon conjugate according to claim 4, wherein the spacer is a polypeptide.
6. An interferon conjugate according to any one of claims 1 to 3, wherein a C-terminal of the region  
20 exhibiting biological activities of interferon- $\beta$  has been linked to an N-terminal of the region exhibiting biological activities of interferon- $\gamma$ , directly or via a spacer.
7. An interferon conjugate according to claim 6,  
25 wherein the interferon- $\beta$  consists of the following amino acid sequence:

MET SER TYR ASN LEU LEU GLY PHE LEU GLN ARG SER SER  
ASN PHE GLN CYS GLN LYS LEU LEU TRP GLN LEU ASN GLY ARG LEU GLU TYR  
CYS LEU LYS ASP ARG MET ASN PHE ASP ILE PRO GLU GLU ILE LYS GLN LEU  
GLN GLN PHE GLN LYS GLU ASP ALA ALA LEU THR ILE TYR GLU MET LEU GLN  
ASN ILE PHE ALA ILE PHE ARG GLN ASP SER SER SER THR GLY TRP ASN GLU  
THR ILE VAL GLU ASN LEU LEU ALA ASN VAL TYR HIS GLN ILE ASN HIS LEU  
LYS THR VAL LEU GLU GLU LYS LEU GLU LYS GLU ASP PHE THR ARG GLY LYS  
LEU MET SER SER LEU HIS LEU LYS ARG TYR TYR GLY ARG ILE LEU HIS TYR  
LEU LYS ALA LYS GLU TYR SER HIS CYS ALA TRP THR ILE VAL ARG VAL GLU  
ILE LEU ARG ASN PHE TYR PHE ILE ASN ARG LEU THR GLY TYR LEU ARG ASN,

and the interferon- $\gamma$  consists of the following amino acid sequence:

GLN ASP PRO TYR VAL LYS GLU ALA GLU ASN LEU LYS LYS  
TYR PHE ASN ALA GLY HIS SER ASP VAL ALA ASP ASN GLY THR LEU PHE LEU  
GLY ILE LEU LYS ASN TRP LYS GLU GLU SER ASP ARG LYS ILE MET GLN SER  
GLN ILE VAL SER PHE TYR PHE LYS LEU PHE LYS ASN PHE LYS ASP ASP GLN  
SER ILE GLN LYS SER VAL GLU THR ILE LYS GLU ASP MET ASN VAL LYS PHE  
PHE ASN SER ASN LYS LYS LYS ARG ASP ASP PHE GLU LYS LEU THR ASN TYR  
SER VAL THR ASP LEU ASN VAL GLN ARG LYS ALA ILE HIS GLU LEU ILE GLN  
VAL MET ALA GLU LEU SER PRO ALA ALA LYS THR GLY LYS ARG LYS ARG SER  
GLN MET LEU PHE ARG GLY ARG ARG ALA SER GLN.

8. An interferon conjugate according to claim 7, wherein a C-terminal of a polypeptide consisting of the following amino acid sequence:

MET SER TYR ASN LEU LEU GLY PHE LEU GLN ARG SER SER  
ASN PHE GLN CYS GLN LYS LEU LEU TRP GLN LEU ASN GLY ARG LEU GLU TYR  
CYS LEU LYS ASP ARG MET ASN PHE ASP ILE PRO GLU GLU ILE LYS GLN LEU  
GLN GLN PHE GLN LYS GLU ASP ALA ALA LEU THR ILE TYR GLU MET LEU GLN  
ASN ILE PHE ALA ILE PHE ARG GLN ASP SER SER SER THR GLY TRP ASN GLU  
THR ILE VAL GLU ASN LEU LEU ALA ASN VAL TYR HIS GLN ILE ASN HIS LEU  
LYS THR VAL LEU GLU GLU LYS LEU GLU LYS GLU ASP PHE THR ARG GLY LYS  
LEU MET SER SER LEU HIS LEU LYS ARG TYR TYR GLY ARG ILE LEU HIS TYR  
LEU LYS ALA LYS GLU TYR SER HIS CYS ALA TRP THR ILE VAL ARG VAL GLU  
ILE LEU ARG ASN PHE TYR PHE ILE ASN ARG LEU THR GLY TYR LEU ARG ASN  
has been linked to an N-terminal of a polypeptide consisting of the following amino acid sequence:

GLN ASP PRO TYR VAL LYS GLU ALA GLU ASN LEU LYS LYS  
TYR PHE ASN ALA GLY HIS SER ASP VAL ALA ASP ASN GLY THR LEU PHE LEU  
GLY ILE LEU LYS ASN TRP LYS GLU GLU SER ASP ARG LYS ILE MET GLN SER  
GLN ILE VAL SER PHE TYR PHE LYS LEU PHE LYS ASN PHE LYS ASP ASP GLN  
SER ILE GLN LYS SER VAL GLU THR ILE LYS GLU ASP MET ASN VAL LYS PHE  
PHE ASN SER ASN LYS LYS LYS ARG ASP ASP PHE GLU LYS LEU THR ASN TYR  
SER VAL THR ASP LEU ASN VAL GLN ARG LYS ALA ILE HIS GLU LEU ILE GLN  
VAL MET ALA GLU LEU SER PRO ALA ALA LYS THR GLY LYS ARG LYS ARG SER  
GLN MET LEU PHE ARG GLY ARG ARG ALA SER GLN,  
via a polypeptide as a spacer.

9. An interferon conjugate according to any one

of claims 1 to 3, wherein a C-terminal of the region exhibiting biological activities of interferon- $\gamma$  has been linked to an N-terminal of the region exhibiting biological activities of interferon- $\beta$ , directly or via a spacer.

10. An interferon conjugate according to claim 9, wherein the interferon- $\gamma$  consists of the following amino acid sequence:

GLN ASP PRO TYR VAL LYS GLU ALA GLU ASN LEU LYS LYS  
TYR PHE ASN ALA GLY HIS SER ASP VAL ALA ASP ASN GLY THR LEU PHE LEU  
GLY ILE LEU LYS ASN TRP LYS GLU GLU SER ASP ARG LYS ILE MET GLN SER  
GLN ILE VAL SER PHE TYR PHE LYS LEU PHE LYS ASN PHE LYS ASP ASP GLN  
SER ILE GLN LYS SER VAL GLU THR ILE LYS GLU ASP MET ASN VAL LYS PHE  
PHE ASN SER ASN LYS LYS LYS ARG ASP ASP PHE GLU LYS LEU THR ASN TYR  
SER VAL THR ASP LEU ASN VAL GLN ARG LYS ALA ILE HIS GLU LEU ILE GLN  
VAL MET ALA GLU LEU SER PRO ALA ALA LYS THR GLY LYS ARG LYS ARG SER  
GLN MET LEU PHE ARG GLY ARG ARG ALA SER GLN,

and the interferon- $\beta$  consists of the following amino acid sequence:

MET SER TYR ASN LEU LEU GLY PHE LEU GLN ARG SER SER  
ASN PHE GLN CYS GLN LYS LEU LEU TRP GLN LEU ASN GLY ARG LEU GLU TYR  
CYS LEU LYS ASP ARG MET ASN PHE ASP ILE PRO GLU GLU ILE LYS GLN LEU  
GLN GLN PHE GLN LYS GLU ASP ALA ALA LEU THR ILE TYR GLU MET LEU GLN  
ASN ILE PHE ALA ILE PHE ARG GLN ASP SER SER SER THR GLY TRP ASN GLU  
THR ILE VAL GLU ASN LEU LEU ALA ASN VAL TYR HIS GLN ILE ASN HIS LEU  
LYS THR VAL LEU GLU GLU LYS LEU GLU LYS GLU ASP PHE THR ARG GLY LYS  
LEU MET SER SER LEU HIS LEU LYS ARG TYR TYR GLY ARG ILE LEU HIS TYR  
LEU LYS ALA LYS GLU TYR SER HIS CYS ALA TRP THR ILE VAL ARG VAL GLU  
ILE LEU ARG ASN PHE TYR PHE ILE ASN ARG LEU THR GLY TYR LEU ARG ASN.

11. An interferon conjugate according to claim 10, wherein a C-terminal of a polypeptide consisting of the following amino acid sequence:

GLN ASP PRO TYR VAL LYS GLU ALA GLU ASN LEU LYS LYS  
TYR PHE ASN ALA GLY HIS SER ASP VAL ALA ASP ASN GLY THR LEU PHE LEU  
GLY ILE LEU LYS ASN TRP LYS GLU GLU SER ASP ARG LYS ILE MET GLN SER  
GLN ILE VAL SER PHE TYR PHE LYS LEU PHE LYS ASN PHE LYS ASP ASP GLN  
SER ILE GLN LYS SER VAL GLU THR ILE LYS GLU ASP MET ASN VAL LYS PHE

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PHE ASN SER ASN LYS LYS LYS ARG ASP ASP PHE GLU LYS LEU THR ASN TYR  
SER VAL THR ASP LEU ASN VAL GLN ARG LYS ALA ILE HIS GLU LEU ILE GLN  
VAL MET ALA GLU LEU SER PRO ALA ALA LYS THR GLY LYS ARG LYS ARG SER  
GLN MET LEU PHE ARG GLY ARG ARG ALA SER GLN

has been linked to an N-terminal of a polypeptide  
consisting of the following amino acid sequence:

MET SER TYR ASN LEU LEU GLY PHE LEU GLN ARG SER SER  
ASN PHE GLN CYS GLN LYS LEU LEU TRP GLN LEU ASN GLY ARG LEU GLU TYR  
CYS LEU LYS ASP ARG MET ASN PHE ASP ILE PRO GLU GLU ILE LYS GLN LEU  
GLN GLN PHE GLN LYS GLU ASP ALA ALA LEU THR ILE TYR GLU MET LEU GLN  
ASN ILE PHE ALA ILE PHE ARG GLN ASP SER SER SER THR GLY TRP ASN GLU  
THR ILE VAL GLU ASN LEU LEU ALA ASN VAL TYR HIS GLN ILE ASN HIS LEU  
LYS THR VAL LEU GLU GLU LYS LEU GLU LYS GLU ASP PHE THR ARG GLY LYS  
LEU MET SER SER LEU HIS LEU LYS ARG TYR TYR GLY ARG ILE LEU HIS TYR  
LEU LYS ALA LYS GLU TYR SER HIS CYS ALA TRP THR ILE VAL ARG VAL GLU  
ILE LEU ARG ASN PHE TYR PHE ILE ASN ARG LEU THR GLY TYR LEU ARG ASN,  
via a polypeptide as a spacer

20 12. A deoxyribonucleotide sequence coding for an  
interferon conjugate comprising in a single molecule a  
region exhibiting biological activities of interferon- $\beta$   
and a region exhibiting biological activities of  
interferon- $\gamma$ .

25 13. A recombinant DNA comprising a deoxyribon-  
ucleotide sequence coding for an interferon conjugate  
comprising in a single molecule a region exhibiting  
biological activities of interferon- $\beta$  and a region  
exhibiting biological activities of interferon- $\gamma$ , and a  
deoxyribonucleotide sequence coding for a control region  
for expression of said interferon conjugate, wherein the  
30 latter sequence is upstream from the former sequence.

35 14. A transformant organism transformed with a  
recombinant DNA comprising a deoxyribonucleotide sequence  
coding for an interferon conjugate comprising in a  
single molecule a region exhibiting biological activities  
of interferon- $\beta$  and a region exhibiting biological  
activities of interferon- $\gamma$ , and a deoxyribonucleotide  
sequence coding for a control region for expression of

said interferon conjugat , wherein the latter sequence is upstream from the former s quence.

15. A process for production of an interf r n conjugate comprising in a single molecule a r gion  
5 exhibiting biological activities of interferon- $\beta$  and a region exhibiting biological activities of interferon- $\beta$ , comprising the step:

culturing a transformant organism transformed with a recombinant DNA comprising a deoxyribonucleotide sequence coding for said interferon conjugate  
10 and a deoxyribonucleotide sequence coding for a control region for expression of the interferon conjugate wherein the latter sequence is upstream from the former sequence, to produce the interferon conjugate;

- 15 and recovering said interferon conjugate.

16. A process according to claim 15, wherein E. coli is used as a host.

17. . A process according to claim 11, wherein human cells are used as a host.

- 20 18. A process according to claim 17, wherein the human cells are cells derived from human lung cancer.

19. A process according to claim 18, wherein the cells derived from human lung cancer are selected from the group consisting of PC8 and PC12 cell lines.



Fig. 1

MET SER TYR ASN

LEU LEU GLY PHE LEU GIN ARG SER SER ASN PHE GIN CYS GIN LYS LEU LEU TRP GIN LEU ASN GLY ARG LEU GLU

TYR CYS LEU LYS ASP ARG MET ASN PHE ASP ILE PRO GLU GLU ILE LYS GIN LEU GIN PHE GIN LYS GLU ASP

ALA ALA LEU THR ILE TYR GLU MET LEU GIN ASN ILE PHE ALA ILE PHE ARG GIN ASP SER SER THR GLY TRP

ASN GLU THR ILE VAL GLU ASN LEU LEU ALA ASN VAL TYR HIS GIN ILE ASN HIS LEU LYS THR VAL LEU GLU GLU

LYS LEU GLU LYS GLU ASP PHE THR ARG GLY LYS LEU MET SER SER LEU HIS LEU LYS ARG TYR TYR GLY ARG ILE

LEU HIS TYR LEU LYS ALA LYS GLU TYR SER HIS CYS ALA TRP THR ILE VAL ARG VAL GLU ILE LEU ARG ASN PHE

TYR PHE ILE ASN ARG LEU THR GLY TYR LEU ARG ASN

Fig. 2

GLY ASP PRO TYR VAL LYS GLU ALA GLU ASN LEU LYS LYS TYR PHE ASN ALA GLY HIS SER ASP VAL  
ALA ASP ASN GLY THR LEU PHE LEU GLY ILE LEU LYS ASN TRP LYS GLU GLU SER ASP ARG LYS ILE MET GIN SER  
GIN ILE VAL SER PHE TYR PHE LYS LEU PHE LYS ASN PHE LYS ASP ASP GIN SER ILE GIN LYS SER VAL GLU THR  
ILE LYS GLU ASP MET ASN VAL LYS PHE PHE ASN SER ASN LYS LYS LYS ARG ASP PHE GLU LYS LEU THR ASN  
TYR SER VAL THR ASP LEU ASN VAL GIN ARG LYS ALA ILE HIS GLU LEU ILE GIN VAL MET ALA GLU LEU SER PRO  
ALA ALA LYS THR GLY LYS ARG LYS ARG SER GIN MET LEU PHE ARG GLY ARG ARG ALA SER GIN

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Fig. 3

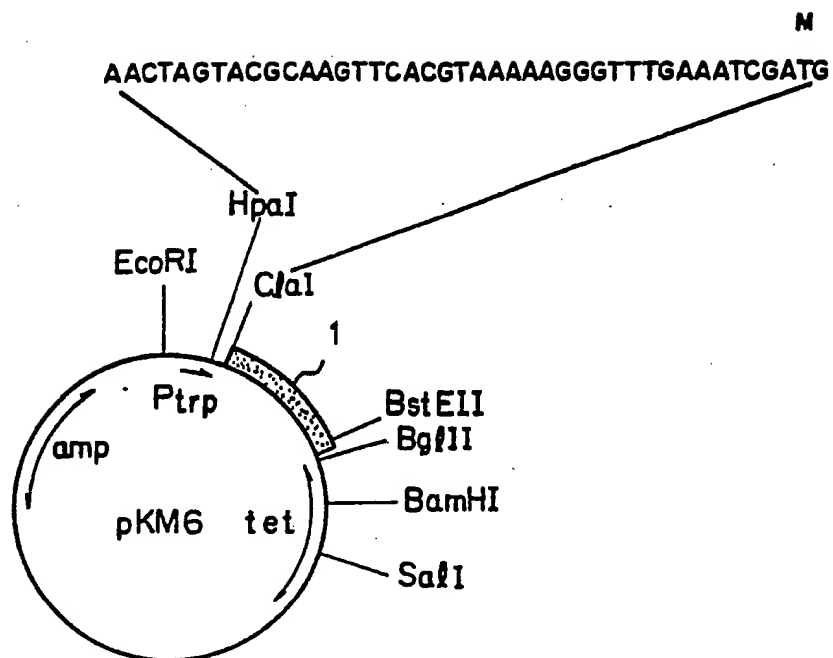


Fig. 4

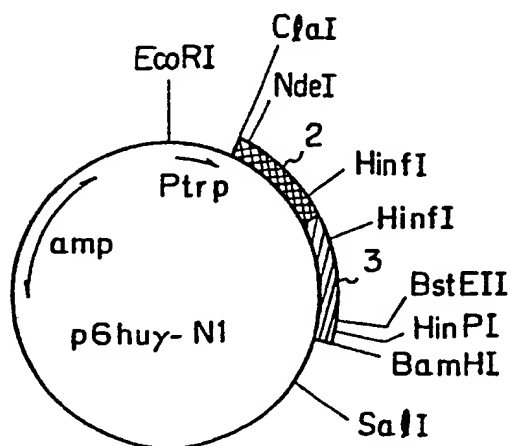


Fig. 5

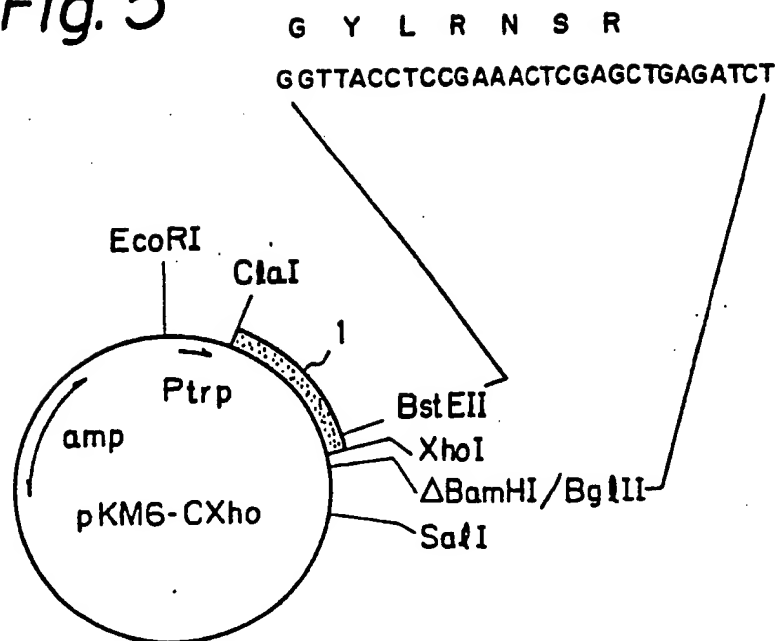


Fig. 6

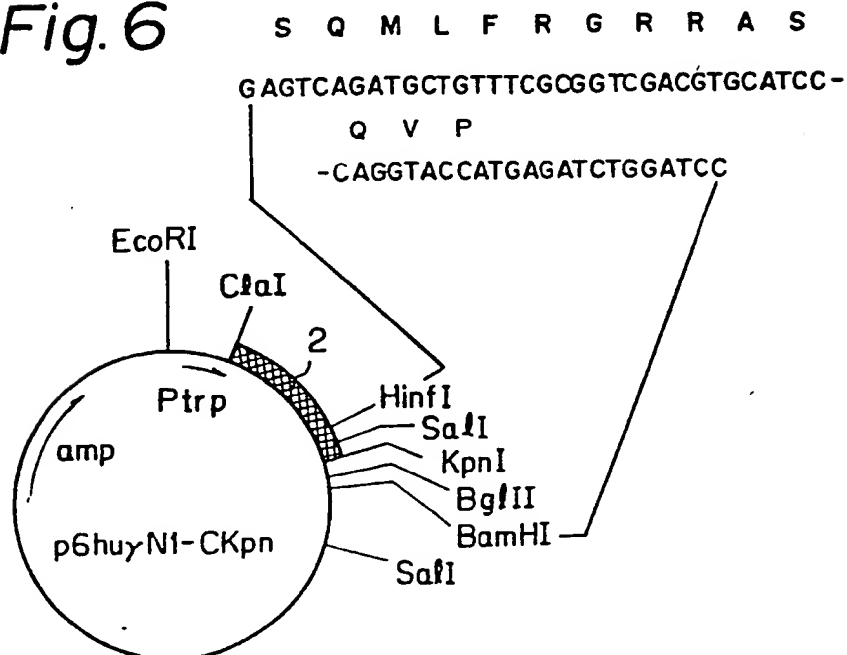


Fig. 7

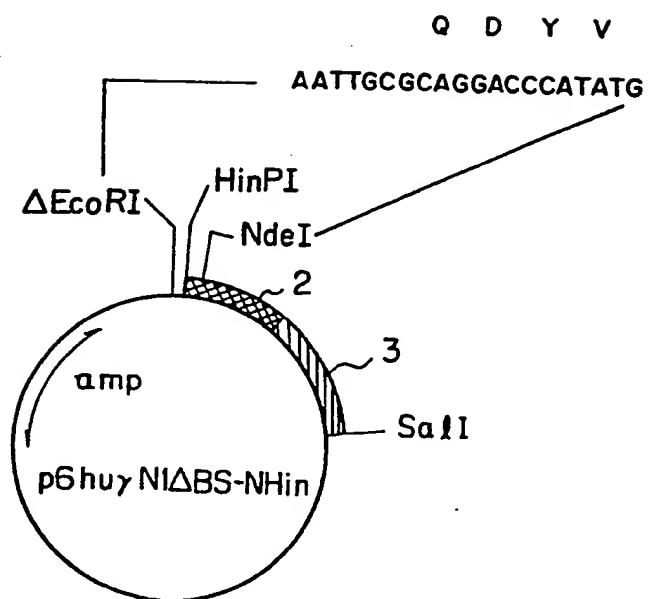


Fig. 8

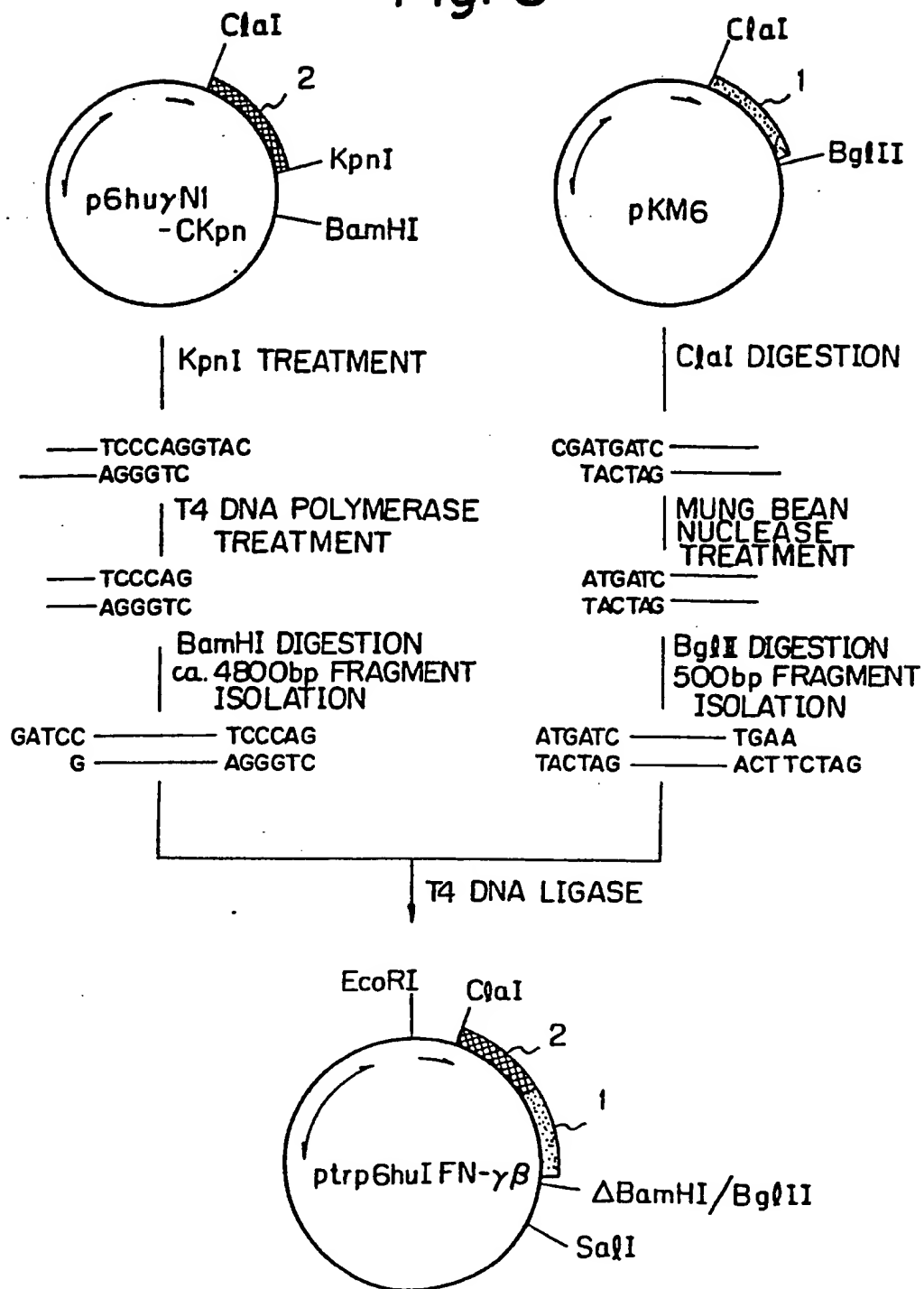


Fig. 9

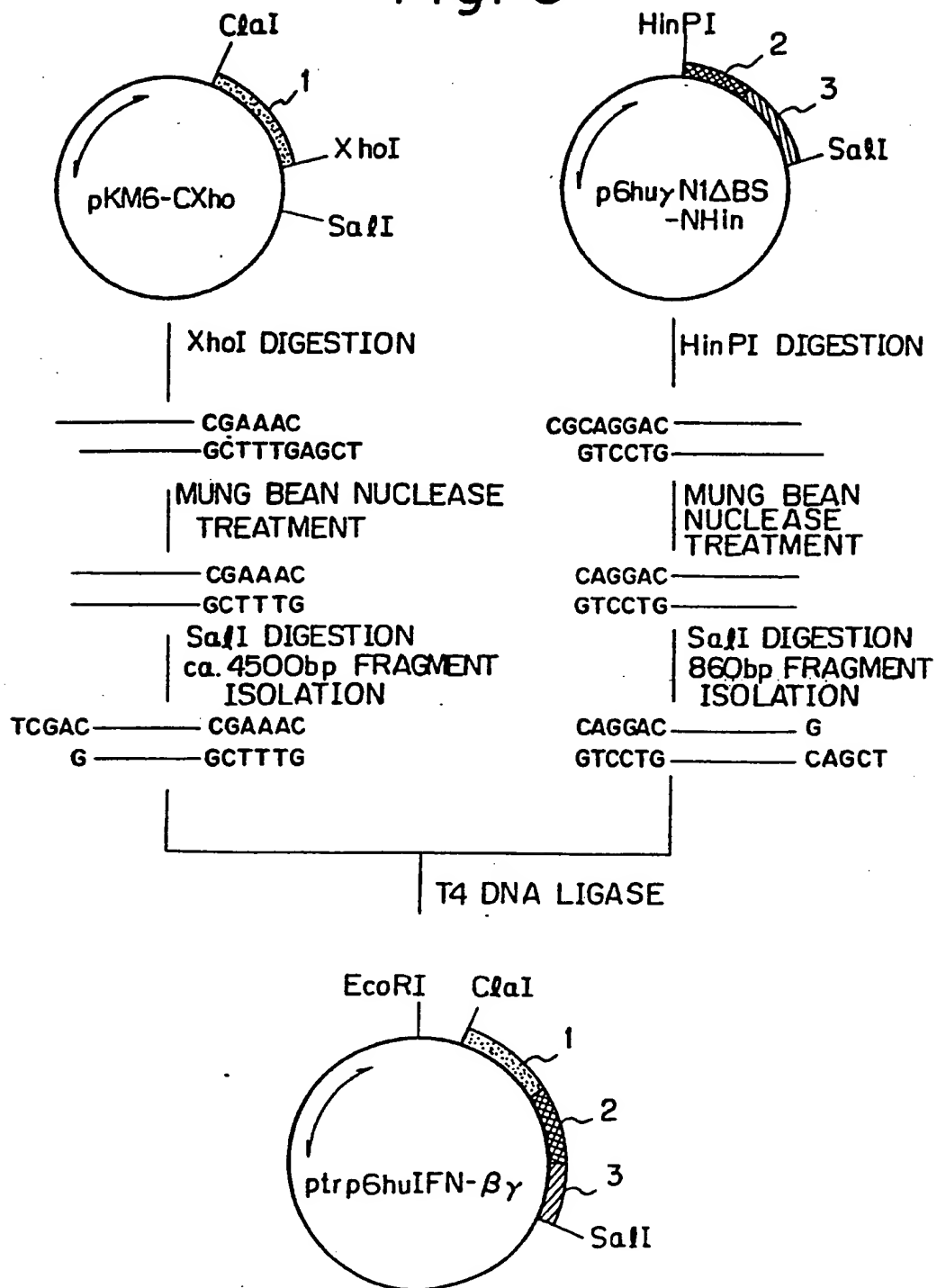
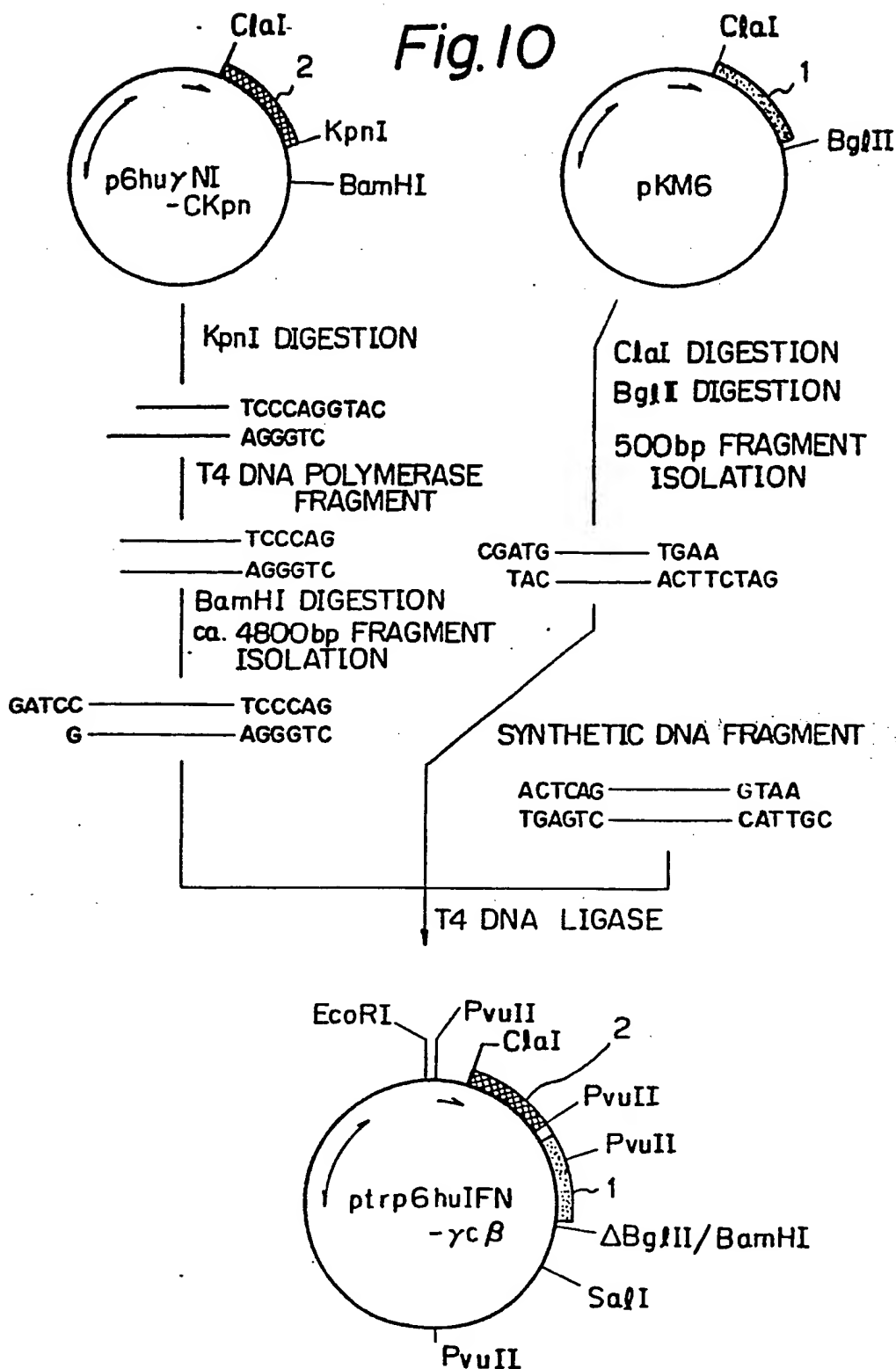


Fig. 10





*Fig. 11*

T Q L G Q P K A A K S V T

ACTCAGCTGGGCCAGCCGAAAGCTGCTAAGTCGGTAA

TGAGTCGACCCGGTCCGGCTTTCGAGCATTTCAGCCATTGC

PvuII

Fig. 12

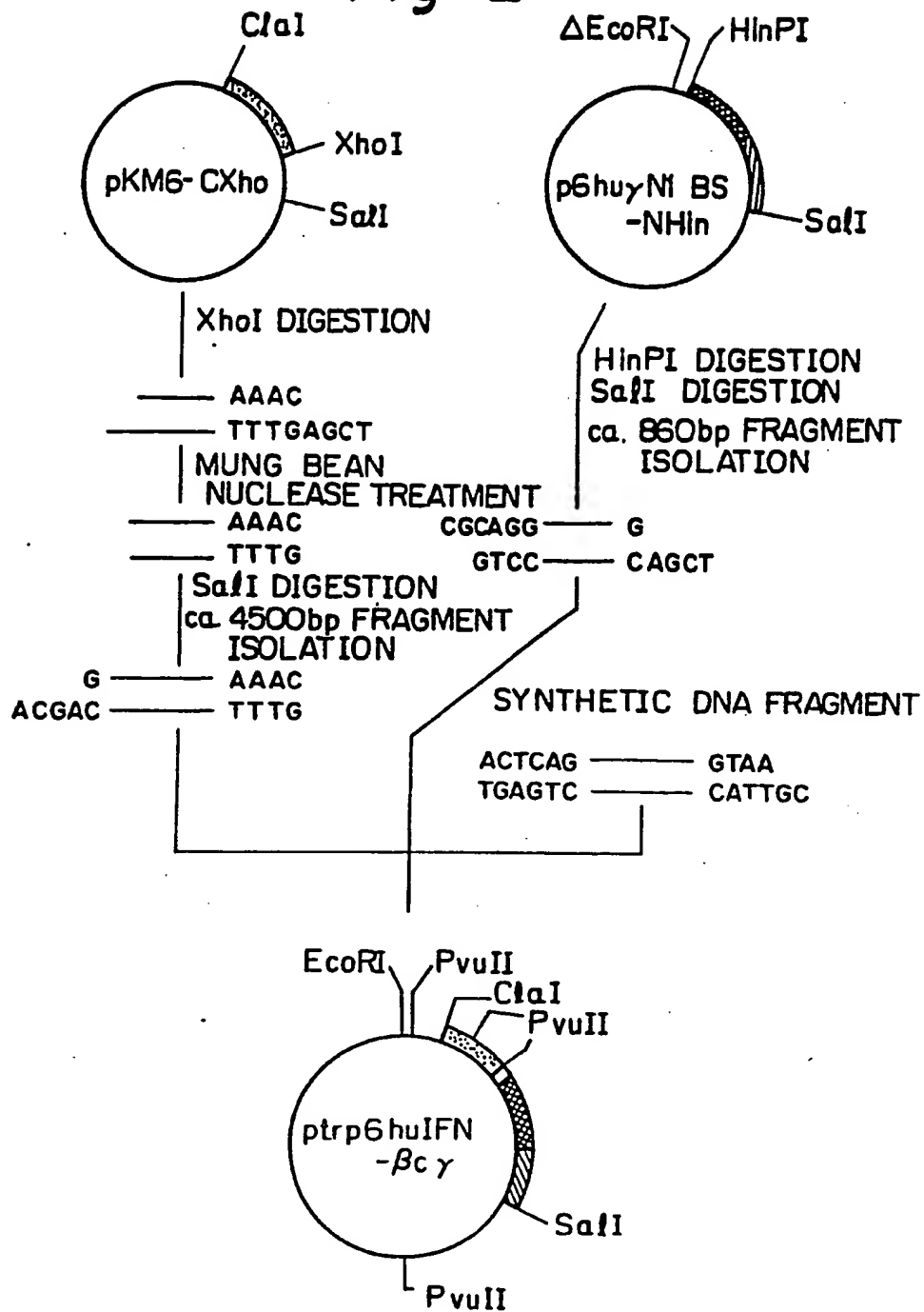


Fig. 13

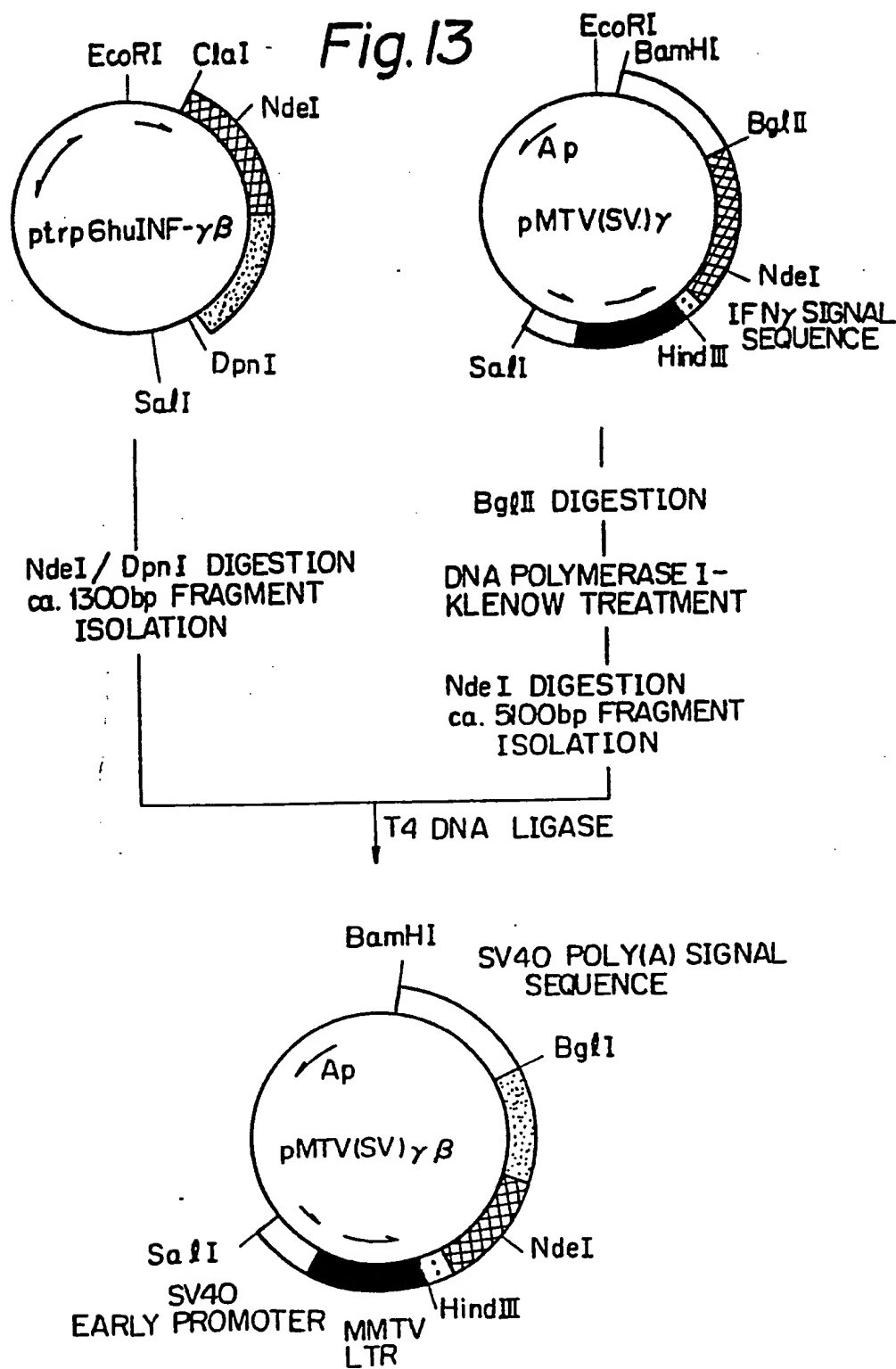


Fig. 14

